

Testicular Histology, Gonadal and Extragonadal Sperm Reserve of Sheep (rams) Fed Tigernut (*Cyperus esculentum*) Chaff and Cowpea (*Vigna unguiculata* L. Walp) Husk in a Mixed Ration.

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Target audience: small ruminant farmers, animal reproductive physiologists, nutritionists.

Abstract

Gonadal, extra-gonadal sperm reserves and testicular histology of Yankasa rams fed tiger nut chaff and cowpea husk in a mixed ration were studied. Thirty growing and apparently healthy Yankasa rams were randomly allotted five dietary treatments with six animals per treatment. Treatments compared were T₁ (Cowpea husk 40% + Tiger nuts chaff 0%), T₂ (Cowpea husk 30% + Tiger nuts chaff 10%), T₃ (Cowpea husk 20% + Tiger nuts chaff 20%), T₄ (Cowpea husk 10% + Tiger nuts chaff 30%), and T₅ (Cowpea husk 0% + Tiger nuts chaff 40%). The testicular morphometry and gonadal and extra-gonadal sperm reserves showed significant differences ($p < 0.05$) in all the parameters observed. For testicular morphometry, the weight and length of the right testes for T₁ were highest (180 g) and (13 cm) and least for T₅ (38 g) and (7cm) respectively. A similar trend was observed for the weight and length of the left testes as 175 g and 12cm, and 39 g and 7cm respectively. Based on the result of this investigation, testicular morphometry, gonadal, and extra-gonadal sperm reserves of rams fed 40% cowpea husk and 0% tiger nuts chaff mixed ration were of the best value. However, semen from animals fed a mixed ration of cowpea husk should be further investigated to determine the impact on the fertility of breeder rams.

Keywords: Testicular histology, gonadal and extra-gonadal sperm reserve, tiger nut, cowpea husk

Description of Problem

Nutrition is a major factor in the effectiveness of reproductive function. It may affect the efficiency of related hormone production and the growth of reproductive

organs. Studies have demonstrated that luteinizing hormone (LH) secretion during early gonadotropin rise is elevated and can be sustained for a longer period when pubertal cattle are fed with improved nutrition during

calf-hood (1). There is a serious shortage in the supply of conventional feed ingredients such as maize, and groundnut cake for a concentrated diet for ruminants. In addition, the production of grains in developing countries is mostly for human consumption, leading to very high prices for the ingredients (2).

Supplementation with unconventional feeds is therefore helpful to reduce the harsh feed scarcity that occurs mostly during the dry season. This to a very large extent will help forestall the consequences of inadequate feed supply experienced during such periods. The utilization of unconventional feed materials that are viable, cheap, and not in high demand by humans (3) is of utmost importance in animal production.

Tiger nut (*Cyperus esculentus*) is one of the underutilized crops. Tiger nut is cultivated for its nutritive, edible nut and has a high content of soluble glucose of about 21% (4). Tiger nut has been reported to have a lot of medicinal benefits (5). Tiger nut is rich in minerals such as potassium 267.18 mg, phosphorus 158.86 mg, calcium 43.36 mg, magnesium 118.14 mg; sodium 17.02 mg; copper 0.54 mg; iron 2.82 mg, and zinc 1.39 mg (6). Tiger nut chaff is obtained after milk extraction from the nut and is discarded as waste. The tiger nut chaff is reported to have a high energy content (7). Despite the numerous nutritional, and health benefits, potentials, and prospects, tiger nut residue as a nutrient source for livestock has not been studied extensively.

Cowpea husk is another unconventional feedstuff. Cowpea (*Vigna unguiculata*) is one of the most popular legume grains in

Africa. It is a member of the Vigna (peas and beans) genus. *Unguiculata* Latin for "with a small claw", which reflects the small stalks on the flower petals (8). Most cowpeas are grown on the African continent, particularly in Nigeria and Niger, which account for 66% of world cowpea production (6). Cowpea husk: a by-product of cowpea production, is obtained after the seeds have been removed by threshing the pods and winnowing out the husks to collect the seeds. The husk is most of the time left on the farm to rot or they are burnt off. Cowpea husk contains 95.49% DM, 12.79% CP, 3.40% CF, 7.00% EE, 10.70% Ash and 41.63% NFE (9). Cowpea husk is characterized by a high level of crude fiber (about 31%) and a relatively low level of protein (12-13%) (10). Cowpea husk has been used in the diets of sheep (11) without adverse effects on the animals. Feeding a mix ration of cowpea husk and tiger nuts chaff is to ascertain if the mix has any deleterious effect on some reproductive indices of rams. For increasing and long-term improvements in animal productivity and profitability, effective reproduction is crucial (12). Examining testicular morphometry is important to forecast not only sperm production but also the storage of sperm. Knowledge about gonadal and extragonadal sperm reserves seems essential for a careful assessment of male fertility (13). Extragonadal sperm reserves (ESR) represent sperm stored in the caput, corpus, and cauda epididymis, and the number of spermatozoa stored in the epididymis has been said to be related to sperm production by the testes (14). This study was designed to investigate the testicular morphometry,

gonadal and epididymal sperm reserves and histology of the testes of Yankasa rams fed a mixed ration containing cowpea husk and tiger nuts chaff.

Materials and Methods

Experimental site

The study was conducted at the small ruminant experimental unit of the National Open University of Nigeria (NOUN), Kaduna farm located at Latitude 10.61362 N 10°36'47.692"

Longitude 7.46852 E 7°28'23.667" at an altitude of 612.65 meters. The climate is tropical as classified by Koppen. The wet season lasts from April through to mid-October with a peak in August; while the dry season extends from mid-October of one calendar year to April the next year (15). The spatial and temporal distribution varies, decreasing from an average of about 1,203 mm. The highest average air temperature occurs in April (28.9°C) and the lowest in December (22.9°C) through January (23.1°C) (16). The mean atmospheric relative humidity ranges between 70% – 90% and 25% – 30 % for the rainy and dry seasons respectively (15).

Experimental Animals and Diet

Thirty (30) growing Yankasa rams aged six to eight months with an initial weight of 15.50 ± 0.67 kg were used for this study. The animals were purchased from an open livestock market in Makarfi LGA of Kaduna State. The pens and the surroundings were cleaned and disinfected with a strong antiseptic (Morigad) two weeks before the arrival of the animals. Upon arrival, the animals were

administered a prophylactic treatment consisting of a long-acting antibiotic (Oxytetracycline 20% LA at 1.0 ml/10kg body weight) intramuscularly, and Ivomec injection at 200 µg/kg (0.3ml/10kg) to control endo and ectoparasites (subcutaneous). The animals were housed in their cages and allowed to adapt to the environment for 14 days before the commencement of the experiment. The cowpea husks used for the study were from the previous harvest from nearby farms that have been allowed to dry in the sun while the tiger nuts chaff used was obtained from a local processor who produces local drink, and the chaff was dried under shade.

Experimental Design and Treatment

The experimental animals were allocated to five dietary treatments with six animals per treatment in a complete randomized design:

T₁ = Cowpea husk 40% + Tiger nuts chaff 0%

T₂ = Cowpea husk 30% + Tiger nuts chaff 10%

T₃ = Cowpea husk 20% + Tiger nuts chaff 20%

T₄ = Cowpea husk 10% + Tiger nuts chaff 30%

T₅ = Cowpea husk 0% + Tiger nuts chaff 40%

Feeding and Management

The study comprised 14 days of the feed adaptation period and 13 weeks (91 days) of the measurement period. Feed was weighed and fed twice daily at 8 a.m. and 2 p.m. Water was provided *ad libitum*. The quantity of feed provided, and the residue of the previous day were weighed to determine the feed intake of each animal. The goats were weighed at the

beginning of the experiment and at weekly intervals in the morning before feeding.

Chemical analysis

The feed samples of experimental diets in triplicates were collected and dried in an air-draft oven at 60°C for 96 hours, ground separately to pass through a 1 mm sieve in a Wiley mill and sampled for chemical analysis using the standard methods of the Association of Official Analytical Chemists (17). Dry matter was determined by drying at 100°C for 24 hours, ash concentration was determined after ignition at 550°C for 4 hours in a muffle furnace and used to calculate organic matter (OM). Fiber fraction analysis was done by the methods of (18). Hemicellulose and cellulose were estimated as differences between neutral detergent fiber (NDF) and acid detergent fiber (ADF) and lignin, respectively.

Statistical Analysis

Data collected were subjected to analysis of variance (ANOVA) in a completely randomized design using the SAS software (19) where significant differences were observed, the means were separated using the Duncan Multiple Range Test (DMRT). Polynomial (linear, quadratic, and cubic)

contrasts were used to determine responses to varying levels of the mixed ration of cowpea husk and tiger nuts chaff.

Determination of Gonadal and Extra Gonadal Sperm Reserves

At the end of the experiment, three rams from each treatment were orchidectomised and the testes and epididymis were used to determine the gonadal and extragonadal (epididymal) sperm reserves, respectively, as described by (20) and (21). Six (6) testes per group were harvested and used for gonadal sperm reserves. Right, and left testes were labelled and appropriately used to determine the gonadal sperm reserves. The length and weight of each testis were determined using a measuring tape and digital weighing balance, respectively. The tunica albuginea was removed from each testis, thereafter, the left and right testes were homogenized separately using a high-speed blender operated for two minutes in 50 ml normal saline that contained 100 IU/ml of Sodium Penicillin G and 1.0 g/ml Streptomycin. The homogenate volume was measured after rinsing the blender with 20 ml of effluent. 5 ml of the homogenate was further diluted with 80 ml of normal saline in a conical flask. The diluted testicular homogenate sample

Table 1: Composition of the experimental diets (%)

Ingredient	Treatment Diets				
	T ₁	T ₂	T ₃	T ₄	T ₅
Tiger nut	0.00	10.00	20.00	30.00	40.00
Cowpea husk	40.00	30.00	20.00	10.00	0.00
Sorghum stover	25.00	25.00	25.00	25.00	25.00
Wheat bran	24.00	24.00	24.00	24.00	24.00
Groundnut cake	10.00	10.00	10.00	10.00	10.00
Salt	0.50	0.50	0.50	0.50	0.50
Premix	0.50	0.50	0.50	0.50	0.50
Total	100.00	100.00	100.00	100.00	100.00

was stored overnight at 5°C and filtered through a gauze then filtrate volume was measured, and sperm concentration was determined using a hemocytometer as described by (20). The *caput*, *corpus*, and *cauda* epididymis were separated using sharp scissors, minced separately in 20 ml of saline, stored overnight at 5°C, and filtered through gauze, then the filtrate volume was measured. One milliliter (ml) of the filtrate was placed in a test tube and further diluted with 2 ml of normal saline. The epididymal sperm reserves were determined using a hemocytometer. Sperm reserves were expressed in billions using the procedures of (22) and (23). Testicular and epididymal weight in grams (g) was obtained using an electronic weighing scale while length was measured using a meter rule in centimeters (cm).

Results

Chemical composition of experimental diets

The chemical composition of the experimental diets is shown in Table 2. From the Table, the dry matter content of the experimental diets ranged from 895.7 g kg⁻¹ DM in T₄ (10% Cowpea husk and 30% Tiger nuts chaff ration mix) to 920.1 g kg⁻¹ DM in T₃ (20% Cowpea husk and 20% Tiger nuts chaff ration mix). T₁ was observed to have the lowest ash content compared to other treatments. The pattern of differences (p<0.05) for lignin and NDF were similar with the highest values (122 g kg⁻¹ DM and 453.7 g kg⁻¹ DM respectively) recorded in T₁ and lowest values (113 g kg⁻¹ DM and 420.6 g kg⁻¹ DM respectively) in T₅. ADF was

significantly different with T₁ recording the highest value of 265.1 g kg⁻¹ DM and lowest in T₅, 232.0 g kg⁻¹ DM. Crude protein values for dietary treatments were significant (p <0.05) with T₁ higher (155.1 g kg⁻¹ DM), lowest in T₅ (145.1 g kg⁻¹ DM), and 150.3 g kg⁻¹ DM, 149.2 g kg⁻¹ DM, 148.0 g kg⁻¹ DM respectively compared to other treatments. A similar trend was observed with the energy content of feeds with T₁ having the highest energy of 10.68MJ and the lowest for T₅ (10.13MJ).

Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on testicular morphometry of Yankasa rams

The result of Tiger nuts chaff and cowpea husk on the testicular morphometry of Yankasa rams is shown in Table 3. Significant differences (p <0.05) were observed among treatments. T₁ had the highest values for testicular weight, and length of the right and left testes. The weight of the right testes for T₁ was highest (180 g) and least for T₅ (38 g). A similar trend was observed for the weight of the left testes as 175 g and 39 g for T₁ and T₅, respectively. The length for the right testes was highest for T₁ (13 cm) and lowest for T₅ (7 cm), also the length for the left testicle was highest for T₁ (12 cm) and lowest for T₅ (7 cm).

Effect of Tiger Nuts Chaff and Cowpea Husk in a mixed ration on Gonadal sperm reserves (x10⁹) of Yankasa rams

There were significant cubic effects (p<0.005, p<0.044) on the gonadal sperm reserve observed among treatments for both the left and right testes as presented in Table

Table 2: Chemical Composition of experimental diets on dry matter (DM) (g/kg DM)

Parameter	T ₁	T ₂	T ₃	T ₄	T ₅	SEM
DM	904.70 ^{ab}	908.00 ^a	920.10 ^a	895.70 ^b	910.10 ^a	1.80
CP	155.10 ^a	150.30 ^{ab}	149.20 ^{ab}	148.00 ^{ab}	145.10 ^b	0.70
EE	41.00 ^a	39.40 ^a	36.70 ^{ab}	35.40 ^{ab}	34.00 ^b	0.60
Ash	68.50 ^b	71.10 ^a	71.80 ^a	72.60 ^a	73.30 ^a	0.40
ADF	265.10 ^a	244.70 ^b	240.60 ^{bc}	235.90 ^{bc}	232.00 ^c	2.60
NDF	453.70 ^a	433.60 ^b	429.00 ^b	422.00 ^b	420.60 ^b	2.70
Lignin	122.00 ^a	117.70 ^b	115.00 ^b	114.20 ^b	113.30 ^b	0.70
Energy (MJ)	10.68 ^a	10.44 ^{ab}	10.26 ^b	10.14 ^c	10.13 ^c	0.05

a,b,c means in the same row with different superscripts are significantly ($p < 0.05$) different, DM=Dry matter, CP=Crude Protein; EE=Ether Extract; ADF=Acid Detergent Fiber; NDF=Neutral Detergent Fiber; SEM=Standard error of mean; T₁ = 40% Cowpea husk, 0% Tiger nut residue; T₂ = 30% Cowpea husk, 10% Tiger nut residue; T₃ = 20% Cowpea husk, 20% Tiger nut residue; T₄ = 10% Cowpea husk, 30% Tiger nut residue; T₅ = 0% Cowpea husk, 40% Tiger nut residue

Table 3: Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on testicular morphometry of Yankasa rams

Parameter	Treatments					P-value			
	T ₁	T ₂	T ₃	T ₄	T ₅	SEM	Linear	Quadratic	Cubic
Right testicular length (cm)	13.00 ^a	10.00 ^b	7.00 ^c	8.00 ^c	7.00 ^c	1.14	0.613	0.910	0.702
Left testicular length (cm)	12.00 ^a	9.00 ^b	6.00 ^c	7.00 ^c	7.00 ^c	1.07	0.252	0.946	0.853
Right testicular weight (g)	180.00 ^a	100.00 ^b	40.00 ^c	40.00 ^c	38.00 ^c	27.72	0.691	0.413	0.425
Left testicular weight (g)	175.00 ^a	150.00 ^a	35.00 ^b	40.00 ^b	39.00 ^b	30.76	0.215	0.731	0.287

a,b,c means in the same row with different superscripts are significantly ($p < 0.05$) different, SEM=Standard error of mean; T₁ = 40% Cowpea husk, 0% Tiger nut residue; T₂ = 30% Cowpea husk, 10% Tiger nut residue; T₃ = 20% Cowpea husk, 20% Tiger nut residue; T₄ = 10% Cowpea husk, 30% Tiger nut residue; T₅ = 0% Cowpea husk, 40% Tiger nut residue

4. Rams in T₁ had the highest value of 2.84×10^9 sperms/testes and T₅ recorded the lowest value of 1.41×10^9 sperms/testes (right testes), and for the left testes, T₁ had a value of 2.83×10^9 /testes and T₅ with 1.24×10^9 /testes.

Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on Extra gonadal sperm reserves of Yankasa rams

The result of extragonadal sperm reserves of Yankasa rams is shown in Table 5. A significant difference ($p < 0.05$) was observed for both left and right epididymides volume. For caput, T₁ had the highest value

for both left (18×10^6 /ml) and right (21×10^6 /ml) while the lowest was observed in T₃ (7×10^6 /ml and 9×10^6 /ml) for left and right epididymides respectively. For corpus, the value for the left and right epididymides for T₁ was highest (7×10^6 /ml and 6×10^6 /ml), and lowest in T₃ (0×10^6 /ml and 2×10^6 /ml). For cauda, T₁ had higher values for both the left (98×10^6 /ml) and right (103×10^6 /ml) epididymides, and lowest in T₃ (18×10^6 /ml and 21×10^6 /ml) for the left and right epididymides respectively.

Table 4: Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on the volume of Testicular homogenate and filtrate of Yankasa rams (mls)

Parameter	Treatments					SEM	P-value		
	T ₁	T ₂	T ₃	T ₄	T ₅		Linear	Quadratic	Cubic
Right testis homogenate	100.00 ^a	80.00 ^b	35.00 ^c	30.00 ^c	40.00 ^c	13.93	0.117	0.381	0.041
Left testis homogenate	95.00 ^a	70.00 ^a	30.00 ^b	31.00 ^b	40.00 ^b	12.72	0.104	0.047	0.225
Right testis filtrate	185.00 ^a	158.00 ^b	70.00 ^c	60.00 ^c	78.00 ^c	11.96	0.241	0.052	0.623
Left testis filtrate	170.00 ^a	130.00 ^b	65.00 ^c	60.00 ^c	70.00 ^c	10.72	0.154	0.413	0.025

a,b,c means in the same row with different superscripts are significantly ($p < 0.05$) different, SEM=Standard error of mean; T₁ = 40% Cowpea husk, 0% Tiger nut residue; T₂ = 30% Cowpea husk, 10% Tiger nut residue; T₃ = 20% Cowpea husk, 20% Tiger nut residue; T₄ = 10% Cowpea husk, 30% Tiger nut residue; T₅ = 0% Cowpea husk, 40% Tiger nut residue

Table 5: Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on Gonadal sperm reserves (x10⁹)/testes of Yankasa rams

Parameter	Treatments					SEM	P-value		
	T ₁	T ₂	T ₃	T ₄	T ₅		Linear	Quadratic	Cubic
Right testes	2.84 ^a	1.88 ^b	1.24 ^c	2.47 ^c	1.41 ^c	0.35	0.151	0.064	0.009
Left testes	2.83 ^a	1.87 ^a	1.03 ^b	2.03 ^b	1.24 ^b	0.32	0.601	0.126	0.044

a,b,c means in the same row with different superscripts are significantly ($p < 0.05$) different, SEM=Standard error of mean; T₁ = 40% Cowpea husk, 0% Tiger nut residue; T₂ = 30% Cowpea husk, 10% Tiger nut residue; T₃ = 20% Cowpea husk, 20% Tiger nut residue; T₄ = 10% Cowpea husk, 30% Tiger nut residue; T₅ = 0% Cowpea husk, 40% Tiger nut residue

Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on the volumes of Testicular homogenate and filtrate of Yankasa rams

Table 6 presents the results of Tiger nuts chaff and cowpea husk in a mixed ration of diets offered to Yankasa rams. The result indicates quadratic and cubic differences ($p < 0.04$) among the homogenate volumes for both the left and right testes across treatments. For the left testes, T₁ had the highest (95 mls), and lowest in T₃ (30 mls). On the other hand, the homogenate volumes for the right testes were highest for T₁ (100 mls) and lowest observed in T₄ (30 mls). There were quadratic ($p < 0.05$) and cubic ($p < 0.03$) differences observed among the filtrate volumes for the right and left testis respectively. For both the right and left testicular filtrates, the highest values

were observed in T₁ with 185 mls and 170 mls and the lowest values were observed in T₄ with 60 mls for both respectively.

The Photomicrograph of a section of the testis

The photomicrograph of the testes reveals degenerated seminiferous tubules for T₁, T₂, T₃, and T₄ but T₅ as presented in Plates 1 to 10, showed a normal histological appearance with aggregated luminal spermatids, well-outlined seminiferous tubule with remains of Leydig cells in between the interstitial spaces.

Discussion

Chemical composition of the diet

The crude protein content was observed to decrease as the level of cowpea husks

Table 6: Effect of Tiger nuts chaff and Cowpea husk in a mixed ration on Extra Gonadal sperm reserves of Yankasa rams ($\times 10^6$ /mls)

Parameter	Treatments					SEM	P-value		
	T ₁	T ₂	T ₃	T ₄	T ₅		Linear	Quadratic	Cubic
Right caput epididymides	21.00 ^a	19.00 ^a	9.00 ^b	15.00 ^{ab}	12.00 ^b	2.20	0.205	0.496	0.583
Left caput epididymides	18.00 ^a	17.00 ^a	7.00 ^b	14.00 ^{ab}	11.00 ^b	2.01	0.678	0.344	0.231
Right carpus epididymides	6.00 ^a	5.00 ^{ab}	2.00 ^c	4.00 ^b	2.00 ^c	0.15	0.905	0.451	0.152
Left carpus epididymides	7.00 ^a	4.00 ^{ab}	0.00	3.00 ^b	2.00 ^b	0.03	0.292	0.705	0.284
Right cauda epididymides	103.00 ^a	80.00 ^b	21.00 ^c	60.00 ^b	71.00 ^b	7.80	0.413	0.510	0.702
Left cauda epididymides	98.00 ^a	73.00 ^{ab}	18.00 ^c	62.00 ^b	57.00 ^b	4.52	0.252	0.946	0.853

^{a,b,c} means in the same row with different superscripts are significantly ($p < 0.05$) different, SEM=Standard error of the mean; T₁ = 40% Cowpea husk, 0% Tiger nut residue; T₂ = 30% Cowpea husk, 10% Tiger nut residue; T₃ = 20% Cowpea husk, 20% Tiger nut residue; T₄ = 10% Cowpea husk, 30% Tiger nut residue; T₅ = 0% Cowpea husk, 40% Tiger nut residue.

decreased and the level of Tiger nuts chaff increased. The crude protein level in this study fell within the range of 139 g kg⁻¹ DM to 220 g kg⁻¹ DM reported by (24) who used cowpea husks to feed goats but lower than the range of 162.1 – 203.3 g kg⁻¹ DM (25) and 354 g kg⁻¹ DM to 383 g kg⁻¹ DM (26) reported. (27) and (28) reported 102.5 and 137.3 g kg⁻¹ DM which is lower than that obtained from this study. The variation observed between previous reports and results from this study could be attributed to the difference in the composition of diets offered to the experimental animals. The crude protein content in this study is above 7% CP recommended for rumen microbes of tropical livestock by (29) below which performance will be deficient. The NDF and ADF were highest at T₁ (453.7 g kg⁻¹ and 265.1 g kg⁻¹ DM respectively) and lowest at T₅ (420.6 g kg⁻¹ and 232.0 g kg⁻¹ DM) which implies that increasing the inclusion of Tiger nuts chaff decreases and the values reported from this experiment is lower than the range of 527 g kg⁻¹ DM – 540.31 g kg⁻¹ DM for NDF and 321.31 g kg⁻¹ DM – 325 g kg⁻¹ DM for ADF as reported by (25). (30) posited that NDF level of forages beyond 65% can limit

feed intake. The EE obtained from this study fell within the range of 22.10 g kg⁻¹ DM - 42.21 g kg⁻¹ DM reported by (25). The Lignin content reported from this study is higher than that reported by (25) and (26). Differences observed in the composition of the feed are attributed to the ration used as (25) used a mix of soya bean meal and *Gmelina arborea* leaves, (26) used cotton seed cake while a mixed ration of cowpea husk and tiger nuts chaff was used for this study.

Testicular histometry

According to (31) and (32), any physical testicular variables that can be measured are closely associated with the ability of the sperm to fertilize, which can be used as an assessment of sperm quality. The size of the reproductive organ and the animal's physiological health might affect the production of semen. The difference observed in the testicular weight and length among treatments may be because the varying levels of cowpea husk and tiger nut chaff had a significant effect on the testicular morphometry as observed in this study, while the testicular weight range from 38 g to 150 g.

Testicular length in this study ranged from 6 cm to 13cm. This report falls within the range reported by (33) for the testicular weight (98.7 g - 158.6 g) and the testicular length (8.2 cm - 9.3 cm) in Yankasa rams. (26) reported lower values for testicular weights but higher values for testicular lengths. The variation observed could be because different animal species and feed rations were used for the studies. (26) used Red Sokoto bucks and fed cotton seed cake while growing Yankasa rams fed a ration mix of cowpea husks and tiger nuts chaff was used for this study. The most suitable testicular parameter that indirectly leads to improving the reproductive performance of the herd, status of the reproductive organ growth, spermatogenesis, and seminal characteristics is the testicular size (34) and it has a positive and strong correlation with sperm production capacity as reported by (35) and (36). (37) reported that heavy-weight testicles generate more spermatozoa than light-weight testicles, as they possess a higher number of Sertoli cells and an excessive number of sperm. Where rams are raised for meat or other purposes aside from reproduction, varied levels of tiger nut chaff could be fed, however, for reproductive purposes, rams fed 40% cowpea husks, and 0% tiger nuts will have optimum performance as they will be sexually more viable.

Testicular homogenate and filtrate

There were significant variations observed in this study on the testicular homogenate and filtrate which does not agree with the findings of (26) and (38) who reported little variation and differences between

homogenate and filtrate volumes? respectively in Red Sokoto bucks fed cotton seed cake. Yankasa rams were used for this study which could have resulted in the observed differences from previous findings because of the variation in the testicular size, shape, and mass. Additionally, variation in testicular homogenate and filtrate volume could also arise from the tissue and its water absorption rate. There is scanty information about the effect of cowpea husk and tiger nut chaff on the testicular homogenate and filtrate of ruminants. Homogenate and filtrate volumes are important indicators of testicular and epididymal sperm reserves in rams, reflecting sperm production, storage, and maturation. Changes in these volumes can impact sperm quality, concentration, and output, ultimately influencing fertility.

Gonadal sperm reserves

(26) and (39) reported lower values for left and right testes (2.76) compared to the values obtained from this study, which could be because Yankasa rams fed cowpea husk, and tiger nuts chaff was used in this study while (26) and (39) used Red Sokoto and Canarian bucks respectively. The results from this study suggest that rams fed 40% cowpea husks had higher spermatogenic action compared to the rams from other treatments. There is a paucity of information on the effect of varying levels of cowpea husks and Tiger nut chaff on gonadal sperm reserves for ruminants.

Extra gonadal sperm reserves

The results from this study do not agree with the report from (26) in Red Sokoto bucks but

concur with the findings of (40) who reported that the epididymides homogenate and filtrate volumes were statistically different ($p < 0.05$) in Akkararnan and Awassi rams. The 'sperm', which is the male reproductive cell, typically takes 49 days (7 weeks) to be generated. Rams' testicular structure is regarded to be a highly accurate predictor of their ability to produce sperm. The relationship between epididymides homogenate and filtrate volumes and spermatogenesis is complex and bidirectional. Changes in volume or composition can impact sperm maturation, concentration, quality, and spermatogenesis regulation, ultimately influencing fertility. The epididymis tail can be palpated to determine sperm reserves. Furthermore, a large, firm tail strongly suggests good storage, while a little, soft tail would suggest the opposite (41). It, therefore, implies that feeding up to 40% of cowpea husk supports increased production of sperm reserves which indicates better reproductive performance.

Histological study

In Plates (1 - 8) there are distorted and degenerated seminiferous tubules in rams fed varying levels of cowpea husk and tiger nuts chaff. The distorted and degenerated seminiferous tubules indicate that there is a limited production of viable sperm cells capable of fertilization which could lead to infertility in extreme cases. Where interstitial edema was observed from the histology of the testes, that could be because of a blockade in spermatogonia differentiation by testosterone thereby indirectly producing

edema as also reported in rats by (42). Inflammatory responses in the testis are very uncommon and are usually seen only in response to seminiferous tubular necrosis or tissue damage sufficient to break down the Sertoli cell tight junctions that make up the blood testis barrier. There are five standard categories of inflammation: acute, suppurative, chronic, chronic-active, and granulomatous (43). Seminiferous tubule vacuoles are in most cases within or between Sertoli cells which is a common early feature of morphologic injury to Sertoli cells that may be seen before any germ cell degeneration (44). Generally, vacuoles often accompany generalized germinal cell degeneration/atrophy (43). Testicular histographs from rams fed a mixed ration of 0% cowpea husk and 40% inclusion of tiger nuts chaff, indicated the existence of well-outlined seminiferous tubules with normal histological appearance of luminal spermatids. The testicular histological changes seen in the present study may be attributed to the low levels of testosterone hormone either directly through the limited number of Leydig cells or indirectly by affecting the central nervous system and release of gonadotropin-releasing hormones (GnRH) from the pituitary as documented by (45). The male gonads' testes contain Leydig cells, which are vital. Together, these cell types support male pubertal sexual traits, oversee hormone balance, and uphold spermatogenesis (46).

Conclusion and Applications

1. The testicular morphometry, gonadal, and extragonadal sperm

reserves gave a better output with a ration mix of 40% cowpea husk and 0% tiger nuts chaff.

2. Semen from animals fed a mixed ration of cowpea husk and tiger nuts chaff should be further investigated to determine the impact of the mixed ration on the fertility of breeder rams.

Acknowledgment

The authors wish to thank the National Open University of Nigeria for funding the research and providing space (Teaching and research farm) where the experiment was conducted.

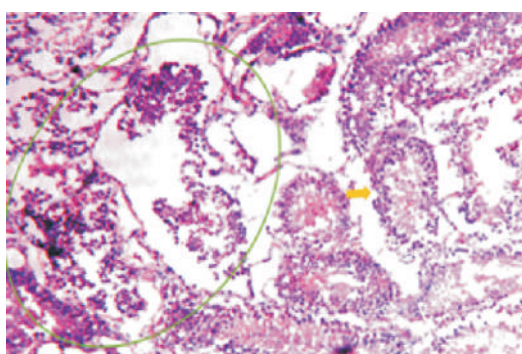


Plate 1: Testicular section (Right) of Yankasa Rams in group T1. The section is showing numerous inflammatory cells (circle) with degenerated and distorted Seminiferous Tubules with what appears to be the remains of spermatids (arrowhead). H&E x250

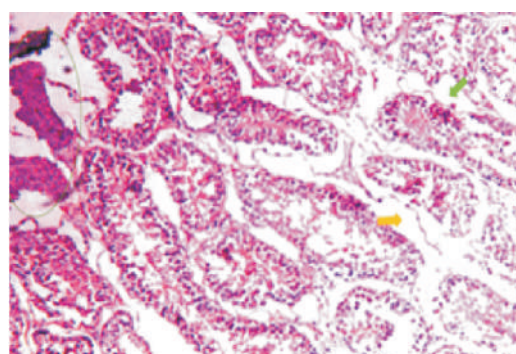


Plate 2: Testicular section (Left) of Yankasa Rams in group T1. The section shows mostly degenerated Seminiferous Tubules (arrowheads) with the complete destruction of some of the spermatozoa (circle). H&E x250

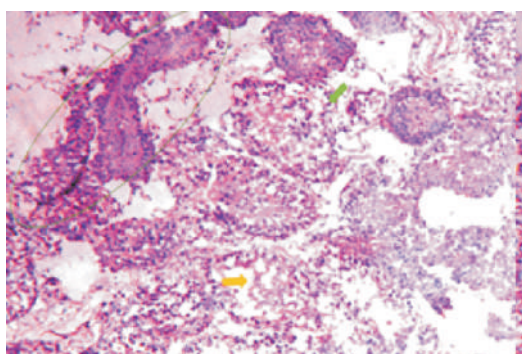


Plate 3: Testicular section (right) of Yankasa Rams in group T2. The section shows mostly degenerated and vacuolated Seminiferous Tubules (arrowheads) with the complete destruction of some of the spermatozoa (circle). H&E x250

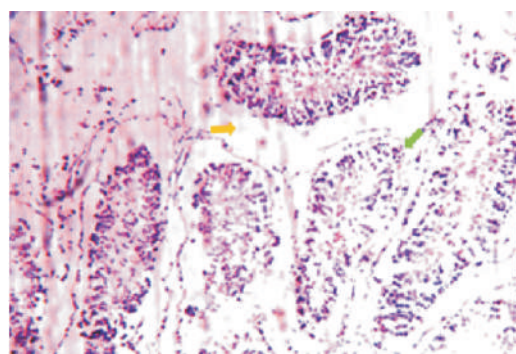


Plate 4: Testicular section (left) of Yankasa Rams in group T2. The section presented ghost Seminiferous Tubules (arrowheads) with the complete destruction of most of the spermatozoa (green arrow) and interstitial

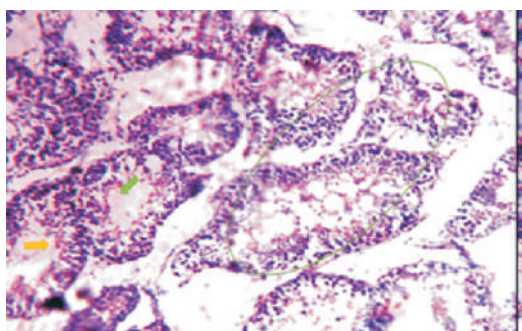


Plate 5: Testicular section (right) of Yankasa Rams in group T3. The section is presenting degenerated (arrowheads) and vacuolated Seminiferous Tubules (circle). H&E x250

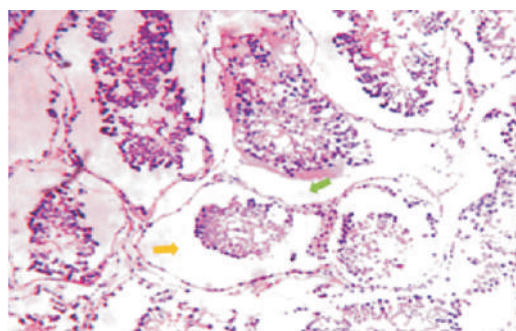


Plate 6: Testicular section (left) of Yankasa Rams in group T3. The section is marked with extensive degenerated Seminiferous Tubules with an appearance of interstitial edema (arrowhead). H&E x250

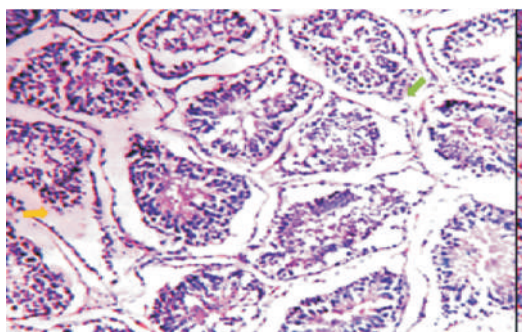


Plate 7: Testicular section (right) of Yankasa Rams in group T4. Most of the Seminiferous Tubules and the spermatozoa appeared degenerated. Note the appearance of interstitial edema (arrowhead). H&E x250

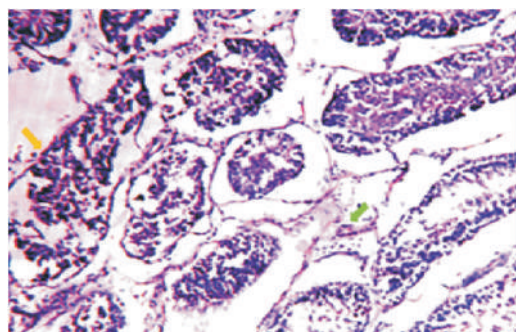


Plate 8: Testicular section (left) of Yankasa Rams in group T4. The section is characterized by degenerated and disoriented spermatozoa and Seminiferous Tubules (arrowheads). The Leydig cells (green arrow) appear preserved. H&E x250

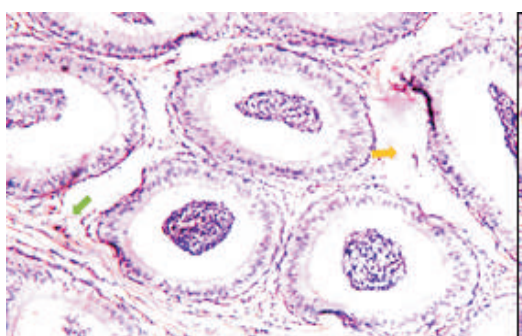


Plate 9: Testicular section (right) of Yankasa Rams in group T5, showing normal histological appearance with aggregated luminal spermatids. The seminiferous tubule is well outlined with remains of Leydig cells (arrow) in between the interstitial spaces. H&E x250

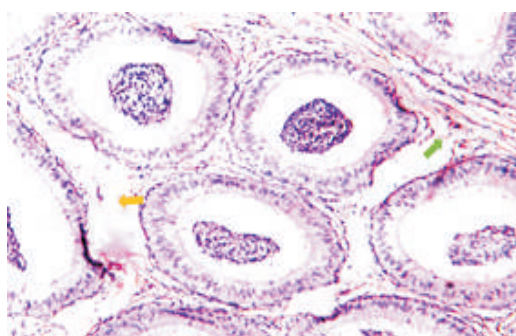


Plate 10: Testicular section (left) of Yankasa Rams in group T5, showing normal histological appearance with aggregated luminal spermatids. The seminiferous tubule is well outlined with remains of Leydig cells (arrow) in between the interstitial spaces. H&E x250

References

1. Cheah, Y. and Yang, W. (2011). Functions of essential nutrition for high-quality spermatogenesis. *Advances in Bioscience and Biotechnology*, 2: 182-197.
2. Cheeke, P.R. and Patton, N. M. (1988). Rabbit production and research in the People's Republic of China. *Journal of Applied Rabbit Research*, 10(2): 68-75.
3. Amaefule, K.U., Nwaokoro, C.C. and Iheukwumere, F.C. (2004). The effect of graded levels of raw pigeon pea (*Cajanus cajan*) seed meal on the performance, nutrient retention, carcass characteristics of weaner rabbits. *Nigerian Journal of Animal Production*, 31(2): 194-199.
4. Bamishaiye, E., Muhammad, N. O. and Bamishaiye, O. M. (2010). Haematological parameters of albino rats fed on tiger nuts (*Cyperus esculentus*) tuber oil meal-based diet. *International Journal of Nutrition and Wellness* (1). DOI: 10.5580/187b.
5. Omode, A. A., Fatoki, O. S. and Olagun, K. A. (1995). Physiochemical Properties of some underexploited and nonconventional oil seeds. *Journal of Agriculture and Food Chemistry*, 43: 2850-2857.
6. FAO (2012). Food and Agricultural Organization of the United Nations. (<https://faostat.fao.org/default.aspx>)
7. Archibong, E. E., Nsa, E. E. and Umoren, U. E. (2018). Nutritional Evaluation of Tigernut (*Cyperus esculentus*) meal as a replacement for maize in broiler diets. *Nigerian Journal of Animal Production*, 45(4): 90-98.
8. Ernest, S. (2009). Top 100 Food Plants. National Research Council, Research Press. pp.104. ISBN 978-0-660-19858-3.
9. Makkar, H. P. S. (2012). Application for the in vitro gas method in the evaluation of feed resources and enhancement of the nutritional value of tannin-rich tree/browse leaves and field evaluation of animal feed supplementation packages. Proceedings of IAEA Cairo, Egypt, pp.23-40.
10. Oluokun, J. A. (2005). Intake, digestion, and nitrogen balance of diets blended with urea-treated and untreated cowpea husk by growing rabbit. *Africa Journal of Biotechnology*, 4(10): 1203–1208.
11. Okoruwa, M. I., Obiku, A. and Agbonlahor, I. (2013). Rumen indices and performance response of West African dwarf (WAD) sheep as influenced by orange with pineapple pulps. *International Journal of Agricultural Science*, 3(11): 862–870.
12. Saleh, B., Mohammed, A. A., Muhammad, A. M., Isah, A. and Zoaka, E. M. (2021). Testicular morphometry, gonadal and extra-gonadal sperm reserves of camel (*Camelus dromedarius*) during the hot dry season in the Sahelian region of Nigeria. *Nigerian Journal of*

- Animal Production*, 48(2): 53-62.
13. Ibrahim, A.A., Aliyu, J., Ashiru, R.M. and Jamilu, M. (2012). Biometric Study of the Reproductive Organs of Three Breeds of Sheep in Nigeria. *International Journal of Morphology*, 30(4):1597-1603.
14. Jindal S. K and Panda J. N. (1980). Epididymal sperm reserves of the goat (*Capra hircus*). *Journal of Reproduction and fertility*, 59: 469-471.
15. Abaje, I. B., Achiebo, P. J. and Matazu, B. M. (2018). Spatio-Temporal Analysis of Rainfall Distribution in Kaduna State. *Ghana Journal of Geography*, 10(1): 1-21.
16. Abaje, I.B., Sawa, B.A., Iguisi, E.O. and Ibrahim, A.A. (2016). Impacts of climate change and adaptation strategies in rural communities of Kaduna State, Nigeria. *Ethiopian Journal of Environmental Studies and Management*, 9(1): 97–108.
17. AOAC (2002). Official Methods of Analysis 16th edition. Association of Official Analytical Chemists, Washington, DC.
18. Van Soest, P.J., Robertson, J.B. and Lewi, B.A. (1991). Methods for dietary fibre, neutral detergent fiber and non-starch polysaccharides in relation to animal nutrition. *Journal of Dairy Science*, 74: 3583-3597.
19. SAS (Statistical Analysis Systems) (2005). *Statistical Package for Analysis* (10.0). Statistical Analysis System Institute, Cary.
20. Rekwot, P. I., Oyedipe, E. O., Akereiola, O. O., Kumi-Diaka, J. and Umoh, J. E. (1987). The effect of intake on pubertal Bunaji and crossbred bull, *Theriogenology*, 28: 427-434.
21. Igboeli, G. and Rakha, A. M. (2007). Seasonal changes in ejaculate characteristics of Angoni (short horn) Zebu bulls. *Journal of Animal Science*, 33: 651-654.
22. Osinowo, O. A., Bale, J. O. and Eduvie, L. O. (1982). Semen quality of Yankasa rams. *Tropical Animal Health and Production*, 14: 181-189.
23. Butswat, I. S. and Zahraddeen, D. (1998). Comparisons of some reproductive parameters in Red Sokoto and Kano Brown breeds of bucks. *Nigerian Journal of Animal Production*, 25(1): 1-5.
24. Olafadehan, O. A., Okunade, S. A., Kholif, A. E., Oludare, T. R., Okoye, G. C., Onwih, E. E., Anaso, E. U., Gero, A. and Shoyombo, A. J. (2022). Urea-amoniated cowpea husk in goat diet: Feed utilization, ruminal fermentation, nutrient digestibility, nitrogen utilization, blood metabolites and growth performance, *Research Square*, (D O I : <https://doi.org/10.21203/rs.3.rs-1753258/v1>).
25. Njidda, A. A., Zikachat, S. S. and Udokpoh, P. R. (2018). Effect of Substituting Soya Bean Meal with *Gmelina arborea* Leaf Meal on Performance and Rumen Fermentation of Red Sokoto Bucks.

- Nigerian Agricultural Journal*, 49(2): 294-301.
26. Nasir, M., Njidda, A. A. and Hassan, A. M. (2014). Testicular histometry, gonadal and extra gonadal Sperm Reserve of Red Sokoto Bucks Fed Cotton Seed Cake. *Journal of Science*, 4(4): 227-232.
27. Abdu, S. B., Hassan, M. R., Jokthan, G. E., Adamu, H. Y., Yashim, S. M. and Yusuf, K. (2012). Effect of varied inclusion levels of gmelina (*Gmelina arborea*) leaf meal on intake, digestibility and nitrogen in red Sokoto bucks fed on sorghum glum based completed diets. *Advances in Agriculture, Sciences and Engineering Research*, 2(2): 79-84. <https://www.ejournal.sedinst.com>.
28. Okafor, E. C., Lakpini, C. A. M. and Fayomi, A. (2012). Dried gmelina (*Gmelina arborea roxb*) leaves as replacement forage to groundnut haulms in the diet of fattening Red Sokoto bucks. *International Journal of Agriculture and Biosciences*, 1(1): 5-10.
29. Minson, D. J. (1990). Forage in Ruminant Nutrition, Academic Press, San Diego, USA, 483–484.
30. Meissner, H. H., Vijj, M. D. and Van Neierkeki, W. A. (1991). Intake and Digestibility of Sheep of Anthephora, panicum Rhode and Smuts Finger Grass Pastures. Proceedings of the 4th International Rangeland Congress, September, Montpellier, France, 1991, 648-649.
31. Akpa, G. N., Ambali, A. L., and Suleiman, I. O. (2012). Relationships between semen cation concentration, semen characteristics, testicular measurement, and body conformation trait in Red Sokoto goat. *Nature and Science*, 11(7): 94-103.
32. Etim, N. A. (2015). Physiological Relationship between Scrotal Morphometric hypoprolactin aemia in the ram plasma gonadotrophin levels, LH and FSH receptors and histology of the testis. *Reproduction and Nutritional Development*, 22: 621-630.
33. Bello, T. K., Rekwot, P. I., Oyelowo, B. B., Hassan, R., Oke-Ebgodo, B. E., Saifullah, U. M., Idris, S. Y. and Abdullahi, M. (2018). Gonadal and Extragonadal Sperm Reserves of Yankasa Rams fed graded levels of Dietary Protein using Cotton seed and Palm Kernel cakes. *International Journal of Agricultural and Biosystems Engineering*, 13: 87-91.
34. Faith, E. A., Sabuwa, A. B. M. and Owoeye, A. O. (2019). Studies on body weight, condition, and scrotal-testicular measurements of rams in Doma and Lafia Metropolis. *EC Veterinary Science*, 4(3): 157-160.
35. Joshi, P. (2022). Nutrition and reproduction in Sheep (Review Article). *Food and Agribusiness Management*, 3(2): 40-44.
36. Mabu, I. M., Mabu, J. M., Anka, B. A.

- and Saheed, Y. (2020). Relationship of testicular biometrics with body weight in rams: A slaughterhouse study in Yobe state, Africa. *Asian Journal of Biotechnology and Genetic Engineering*, 3(3):39-46.
37. Sofi, S. A. (2022). Morphological and biometrical study on adult rams, bucks, and bull's testis in Duhok province. *Journal of Agriculture and Veterinary Services*, 25(1): 38-45.
38. Fadason, S. (2001). Effects of intratesticular injection of chlorohexidine gluconate and cotton seed oil of some reproductive parameters in bucks, MSc Thesis, Ahmadu Bello University, Zaria.
39. Batista, M., Prats, N. Calero, P., Gonzalez, F., Cabrena, F., Medrano, A. and Gracia A. (2002). Semen characteristics and plasma levels of testosterone after vasectomy in bucks. *Reproduction in Domestic Animals*, 37(6):375-378.
40. Gundogan, M. (2006). Some reproductive parameters and seminal plasma constituents in relation to season in Akkararnan and Awassi rams. *Turkish Journal of Veterinary Animal Science*, 30: 95-100.
41. Aqeel, M., AL-Mahmodi, M., Waleed, J., AL-Kelaby, J., Ali, A. and AL-Ramahy, J. (2017). Testis Morpho-biometrical studies of the adult Ram and Buck in AL-Najaf AL-Ashraf Province. *Mirror of Research in Veterinary Sciences and Animals*, 6(3): 32-39.
42. Porter, L. K., Shetty, G. and Meistrich, M. L. (2006). Testicular edema is associated with spermatogonia arrest in irradiated rats. *Endocrinology*, 147(3): 1297-1305.
43. Creasy D., Bube, A., Rijk, E., Kandori, H., Kuwahara, M., Masson, R., Nolte, T., Reams, R., Regan, K., Rehm, S., Rogerson, P. and Whitney, K. (2012). Proliferative and nonproliferative lesions of the rat and mouse male reproductive system. *Toxicology and Pathology*, 40: 40S-121S.
44. Creasy, D. M., Maronpot, R. R. and Giri, K. D. (2014). Testis, seminiferous tubule – vacuolation. National Toxicology Program, U.S. Department of Health and Human Services. Nonneoplastic Lesion Atlas.
45. Kumar, A., Gupte, B. and Shangloo, P. (2021). Effect of Nicotine Smoke on Genitourinary Organs of Male Albino Rats: An Experimental Study. *International Journal of Scientific Research in Dental and Medical Sciences*, 3: 141-146. <http://doi.org/10.30485/IJSRDMS.2021.296503.1177>.
46. Aladamat, N. and Tadi, P. (2021). Histology, Leydig Cells. Stat Pearls Publishing LLC.